

Wantai Hepatitis E Virus Diagnostics

WANTAI HEV-IgG ELISA

Diagnostic Kit for IgG Antibody to Hepatitis E Virus (ELISA)



WE-7296



V. 2016-01 [Eng.]



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Read the package insert carefully and completely before performing the assay. Follow the instructions and do not modify them. Only by strict adherence to these instructions, the erroneous results can be avoided and the optimal performance of WANTAI HEV-IgG ELISA achieved.

INTENDED USE

WANTAI HEV-IgG ELISA is an enzyme-linked immunosorbent assay for the qualitative detection of IgG-class antibodies to hepatitis E virus in human serum or plasma. It is intended to be used as an aid in supplementary diagnosis to acute hepatitis E infection and prevalence studies among the population.

SUMMARY

Hepatitis E virus (HEV) is a non-enveloped, single-stranded RNA virus identified in 1990. Infection with HEV induces acute or sub-clinical liver diseases similar to hepatitis A. HEV infections, endemic and frequently epidemic in developing countries, is seen also in developed countries in a sporadic form with or without a history of traveling to endemic area. The overall case-fatality is 0.5–3%, and much higher (15–25%) among pregnant women. A hypothesis that HEV infection is a zoonosis was presented in 1995. Then a swine HEV and later an avian HEV were identified and sequenced separately in 1997 and 2001. Since then, HEV infection include anti-HEV, viremia and feces excretion of HEV was seen in a wide variety of animals, i.e., swine, rodents, wild monkeys, deer, cow, goats, dogs and chicken in both the developing and developed countries. A recent testimony was reported that the consumption of uncooked deer meat infected with HEV led to acute hepatitis E in human. And HEV genome sequences can be detected in pork livers available in the supermarkets in Japan.

PRINCIPLE OF THE TEST

WANTAI HEV-IgG ELISA employs solid phase, indirect ELISA method for detection of IgG-class antibodies to HEV (anti-HEV) in two-step incubation procedure. Polystyrene microwell strips are pre-coated with HEV recombinant. During the first incubation step, HEV specific antibodies, if present, will be bound to the solid phase pre-coated HEV antigens. The wells are washed to remove unbound serum proteins and then, anti-human IgG antibodies (anti-IgG) conjugated to horseradish peroxidase (HRP-Conjugate) is added. During the second incubation step, these HRP-conjugated antibodies will be bound to any antigen-antibody (IgG) complexes previously formed and the unbound HRP-conjugate is then removed by washing. Chromogen solutions containing Tetramethyl benzidine (TMB) and urea peroxide are added to the wells and in presence of the antigen-antibody-anti-IgG immunocomplex, the colorless Chromogens are hydrolyzed by the bound HRP conjugate to a blue colored product. The blue color turns yellow after stopping the reaction with sulfuric acid. The amount of color intensity can be measured and is proportional to the amount of antibody captured in the wells, and to the specimen respectively. Wells containing specimens negative for HEV-IgG remain colorless.

COMPONENTS



In Vitro Diagnostic Use Only

This kit contains reagents sufficient for testing of maximum of 91 specimens in a test run.

UWELL PLATE

Code 5 (1x96wells)
8x12/12x8-well per plate

CONTROL -

Code 8 (1x0.5ml per vial)
preserv.0.1% ProClin™ 300

CONTROL +

Code 7 (1x0.5ml per vial)
preserv.0.1% ProClin™ 300

HRP CON

Code 6 (1x12ml per vial)
preserv.0.1% ProClin™ 300

DIL SPE

Code 9 (1x12ml per vial)
preserv.0.1% ProClin™ 300

WASH BUF 20X

Code 1 (1x50ml per bottle)
DILUTE BEFORE USE!
detergent Tween-20

CHROM A

Code 2 (1x7ml per vial)

CHROM B

Code 3 (1x7ml per vial)

STOP SOL

Code 4 (1x7ml per vial)

STOP SOLUTION:

Colorless liquid in a white vial with yellow screw cap.
Diluted sulfuric acid solution (0.5M H₂SO₄).
Ready to use as supplied. Once opened, stable for 4 weeks at 2-8°C.

- PLASTIC SEALABLE BAG: For enclosing the strips not in use 1 unit
- PACKAGE INSERT 1 copy
- CARDBOARD PLATE COVER 2 sheets

To cover the plates during incubation and prevent evaporation or contamination of the wells.

MATERIALS REQUIRED BUT NOT PROVIDED

Freshly distilled or deionized water, disposable gloves and timer, appropriate waste containers for potentially contaminated materials, dispensing system and/or pipette, disposable pipette tips, absorbent tissue or clean towel, dry incubator or water bath, 37±1°C, plate reader, single wavelength 450nm or dual wavelength 450/600–650nm, microwell aspiration/wash system.

SPECIMEN COLLECTION, TRANSPORTING AND STORAGE

1. **Specimen Collection:** No special patient's preparation required. Collect the specimen in accordance with the normal laboratory practice. Either fresh serum or plasma specimens can be used with this assay. Blood collected by venipuncture should be allowed to clot naturally and completely – the serum/plasma must be separated from the clot as early as possible as to avoid haemolysis of the RBC. Care should be taken to ensure that the serum specimens are clear and not contaminated by microorganisms. Any visible particulate matters in the specimen should be removed by centrifugation at 3000 RPM (round per minutes) for 20 minutes at room temperature or by filtration.
2. Plasma specimens collected into EDTA, sodium citrate or heparin can be tested, but **highly lipaemic, icteric, or hemolytic specimens should not be used** as they can give false results in the assay. **Do not heat inactivate specimens.** This can cause deterioration of the target analyte. Specimens with visible microbial contamination should never be used.
3. WANTAI HEV-IgG ELISA is intended ONLY for testing of individual serum or plasma specimens. Do not use the assay for testing of cadaver specimens, saliva, urine or other body fluids, or pooled (mixed) blood.
4. **Transportation and Storage:** Store specimens at 2-8°C. Specimens not required for assaying within 1 week should be stored frozen (-20°C or lower). Multiple freeze-thaw cycles should be avoided. For shipment, specimens should be packaged and labeled in accordance with the existing local and international regulations for transportation of clinical specimens and etiological agents.

STORAGE AND STABILITY

The components of the kit will remain stable through the expiration date indicated on the label and package when stored between 2-8°C, do not freeze. To assure maximum performance of WANTAI HEV-IgG ELISA, during storage, protect the reagents from contamination with microorganism or chemicals.

PRECAUTIONS AND SAFETY

TO BE USED ONLY BY QUALIFIED PROFESSIONALS

The ELISA assays are time and temperature sensitive. To avoid incorrect result, **strictly follow the test procedure steps and do not modify them.**

1. Do not exchange reagents from different lots or use reagents from other commercially available kits. The components of the kit are precisely matched for optimal performance of the tests.
2. Make sure that all reagents are within the validity indicated on the kit box and of the same lot. Never use reagents beyond their expiry date stated on labels or boxes.
3. **CAUTION - CRITICAL STEP:** Allow the reagents and specimens to reach room temperature (18-30°C) before use. Shake reagent gently before use. Return at 2-8°C immediately after use.
4. Use only sufficient volume of specimen as indicated in the procedure steps. Failure to do so, may cause low sensitivity of the assay.
5. Do not touch the exterior bottom of the wells; fingerprints or scratches may interfere with the reading. When reading the results, ensure that the plate bottom is dry and there are no air bubbles inside the wells.
6. Never allow the microplate wells to dry after the washing step. Immediately proceed to the next step. Avoid the formation of air bubbles when adding the reagents.
7. Avoid long time interruptions of assay steps. Assure same working conditions for all wells.
8. Calibrate the pipette frequently to assure the accuracy of specimens/reagents dispensing. Use different disposal pipette tips for each specimen and reagents in order to avoid cross-contaminations.
9. Assure that the incubation temperature is 37°C inside the incubator.
10. When adding specimens, do not touch the well's bottom with the pipette tip.
11. When measuring with a plate reader, determine the absorbance at 450nm or at 450/600–650nm.
12. The enzymatic activity of the HRP-conjugate might be affected from dust and reactive chemical and substances like sodium hypochlorite, acids, alkalis etc. Do not perform the assay in the presence of these substances.
13. If using fully automated equipment, during incubation, do not cover the plates with the plate cover. The tapping out of the remainders inside the plate after washing, can also be omitted.
14. All specimens from human origin should be considered as potentially infectious. Strict adherence to GLP (Good Laboratory Practice) regulations can ensure the personal safety.
15. **WARNING:** Materials from human origin may have been used in the preparation of the Negative Control of the kit. These materials have been tested with tests kits with accepted performance and found negative for HBSAg and antibodies to HIV 1/2, HCV, TP. However, there is no analytical method that can assure that infectious agents in the specimens or reagents are completely absent. Therefore, handle reagents and specimens with extreme caution as if capable of transmitting infectious diseases. Bovine derived sera have been used for stabilizing of the positive and negative controls. Bovine serum albumin (BSA) and fetal calf sera (FCS) are derived from animals from BSE/TSE free-geographical areas.
16. Never eat, drink, smoke, or apply cosmetics in the assay laboratory. Never pipette solutions by mouth.
17. Chemical should be handled and disposed of only in accordance with the current GLP (Good Laboratory Practices) and the local or national regulations.
18. The pipette tips, vials, strips and specimen containers should be collected and autoclaved for not less than 2 hours at 121°C or treated with 10% sodium hypochlorite for 30 minutes to decontaminate before any further steps of disposal. Solutions containing sodium hypochlorite should NEVER be autoclaved. Materials Safety Data Sheet (MSDS) available upon request.
19. Some reagents may cause toxicity, irritation, burns or have carcinogenic effect as raw materials. Contact with the skin and the mucosa should be avoided but not limited to the following reagents: Stop solution, the

20. Chromogens, and the Wash buffer.
The Stop solution 0.5M H₂SO₄ is an acid. Use it with appropriate care. Wipe up spills immediately and wash with water if come into contact with the skin or eyes.
21. ProClin™ 300 0.1% used as preservative, can cause sensation of the skin. Wipe up spills immediately or wash with water if come into contact with the skin or eyes.

INDICATIONS OF INSTABILITY DETERIORATION OF THE REAGENT: Values of the Positive or Negative controls, which are out of the indicated quality control range, are indicators of possible deterioration of the reagents and/or operator or equipment errors. In such case, the results should be considered as invalid and the specimens must be retested. In case of constant erroneous results and proven deterioration or instability of the reagents, immediately substitute the reagents with new one or contact Wantai technical support for further assistance.



Warning:
H317, P280, P333+P313, P363
ProClin™ 300



Danger:
H360D, P201, P280, P308+P313
N,N- dimethylformamide

PROCEDURE

Reagents preparation: Allow the reagents to reach room temperature (18-30°C). Check the Wash buffer concentrate for the presence of salt crystals. If crystals have formed, resolubilize by warming at 37°C until crystals dissolve. Dilute the Wash buffer (20X) as indicated in the instructions for washing. Use distilled or deionized water and only clean vessels to dilute the buffer. All other reagents are **READY TO USE AS SUPPLIED**.

- Step 1 Preparation:** Mark three wells as Negative control (e.g. B1, C1, D1), two wells as Positive control (e.g. E1, F1) and one Blank (e.g. A1, neither specimens nor HRP-Conjugate should be added into the Blank well). If the results will be determined by using dual wavelength plate reader, the requirement for use of Blank well could be omitted. Use only number of strips required for the test.
- Step 2 Adding Diluent:** Add 100µl of Specimen Diluent into each well except the Blank.
- Step 3 Adding Specimen:** Add 10µl of Positive control, Negative control, and Specimen into their respective wells except the Blank. **Note: Use a separate disposal pipette tip for each specimen, Negative Control, Positive Control to avoid cross-contamination.** Mix by tapping the plate gently.
- Step 4 Incubating:** Cover the plate with the plate cover and incubate at 37°C for 30 minutes.
- Step 5 Washing:** At the end of the incubation, remove and discard the plate cover. Wash each well 5 times with diluted Wash Buffer. Each time allow the microwells to soak for 30-60 seconds. After the final washing cycle, turn down the plate onto blotting paper or clean towel, and tap it to remove any remainders.
- Step 6 Adding HRP-Conjugate:** Add 100µl of HRP-Conjugate into each well except the Blank.
- Step 7 Incubating:** Cover the plate with the plate cover and incubate at 37°C for 30 minutes.
- Step 8 Washing:** At the end of the incubation, remove and discard the plate cover. Wash each well 5 times with diluted Wash Buffer. Each time allow the microwells to soak for 30-60 seconds. After the final washing cycle, turn down the plate onto blotting paper or clean towel and tap it to remove any remainders.
- Step 9 Coloring:** Add 50µl of Chromogen Solution A and then 50µl of Chromogen Solution B into each well including the Blank, mix gently. Incubate the plate at 37°C for 15 minutes avoiding light. The enzymatic reaction between the Chromogen solutions and the HRP-Conjugate produces blue color in Positive control and HEV IgG positive specimen wells.
- Step 10 Stopping Reaction:** Using a multichannel pipette or manually, add 50µl of Stop Solution into each well and mix gently. Intensive yellow color develops in Positive control and HEV IgG positive specimen wells.
- Step 11 Measuring the Absorbance:** Calibrate the plate reader with the Blank well and read the absorbance at 450nm. If a dual filter instrument is used, set the reference wavelength at 600–650nm. Calculate the Cut-off value and evaluate the results. (**Note:** read the absorbance within 10 minutes after stopping the reaction).

INSTRUCTIONS FOR WASHING

1. A good washing procedure is essential in order to obtain correct and precise analytical data.
2. It is therefore, recommended to use a good quality ELISA microplate washer, maintained at the best level of washing performances. In general, no less than 5 automatic washing cycles of 350-400µl/well are sufficient to avoid false positive reactions and high background.
3. To avoid cross-contaminations of the plate with specimen or HRP-conjugate, after incubation, do not discard the content of the wells but allow the plate washer to aspirate it automatically.
4. Assure that the microplate washer liquid dispensing channels are not blocked or contaminated and sufficient volume of Wash buffer is dispensed each time into the wells.
5. In case of manual washing, we suggest to carry out 5 washing cycles, dispensing 350-400µl/well and aspirating the liquid for 5 times. If poor results (high background) are observed, increase the washing cycles or soaking time per well.
6. In any case, the liquid aspirated out the strips should be treated with a sodium hypochlorite solution at a final concentration of 2.5% for 24 hours, before they are wasted in an appropriate way.
7. The concentrated Wash buffer should be diluted 1 to 20 before use. If less than a whole plate is used, prepare the proportional volume of solution.

QUALITY CONTROL AND CALCULATION OF THE RESULTS

Each microplate should be considered separately when calculating and interpreting the results of the assay, regardless of the number of plates concurrently processed. The results are calculated by relating each specimen absorbance (A) value to the Cut-off value (C.O.) of the plate. If the Cut-off reading is based on single filter plate reader, the results should be calculated by subtracting the Blank well A value from the print report values of specimens and controls. In case the reading is based on dual filter plate reader, do not subtract the Blank well A value from the print report values of specimens and controls.

Calculation of the Cut-off value (C.O.) = Nc + 0.16
(Nc = the mean absorbance value for three negative controls).

Important: If the mean A value of the negative controls is lower than 0.03, take it as 0.03.

Quality control (assay validation): The test results are valid if the Quality Control criteria are fulfilled. It is recommended that each laboratory must establish appropriate quality control system with quality control material similar to or identical with the patient specimen being analyzed.

- The A value of the Blank well, which contains only Chromogen and Stop solution, is < 0.080 at 450nm.
- The A values of the Positive control must be ≥ 0.800 at 450/600–650nm or at 450nm after blanking.
- The A values of the Negative control must be ≤ 0.100 at 450/600–650nm or at 450nm after blanking.

If one of the Negative control A values does not meet the Quality Control criteria, it should be discarded, and the mean value should be calculated by using the remaining two values. If more than one Negative control A values do not meet the Quality Control Range specifications, the test is invalid and must be repeated.

Example:

1. Quality Control

Blank well A value: A1= 0.025 at 450nm (Note: blanking is required only when reading with single filter at 450nm)

Well No.: B1 C1 D1
 Negative control A values after blanking: 0.020 0.012 0.016

Well No.: E1 F1
 Positive control A values after blanking: 2.421 2.369

All control values are within the stated quality control range

2. Calculation of Nc: = $(0.020+0.012+0.016) / 3 = 0.016$ (Nc is lower than 0.03, so take it as 0.03)

3. Calculation of the Cut-off: (C.O.) = $0.03 + 0.16 = 0.19$

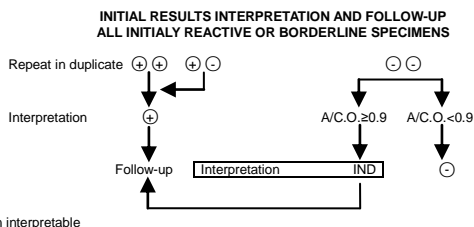
INTERPRETATIONS OF THE RESULTS

Negative Results (A / C.O. < 1): Specimens giving A value less than the Cut-off value are negative for this assay, which indicates that no HEV IgG antibodies have been detected with WANTAI HEV-IgG ELISA, therefore there are no serological indications for infection with HEV.

Positive Results (A / C.O. ≥ 1): Specimens giving A value equal to or greater than the Cut-off value are considered initially reactive, which indicates that HEV IgG antibodies have probably been detected with WANTAI HEV-IgG ELISA. Retesting in duplicate of any initially reactive specimen is recommended. Repeatedly reactive specimens could be considered positive for HEV IgG antibodies and therefore the patient is probably infected with HEV.

Borderline (A / C.O. = 0.9-1.1): Specimens with A value to Cut-off ratio between 0.9 and 1.1 are considered borderline and retesting of these specimens in duplicate is required to confirm the initial results.

Follow-up, confirmation and supplementary testing of any positive specimen with other analytical system (e.g. PCR) is required. Clinical diagnosis should not be established based on a single test result. It should integrate clinical and other laboratory data and findings.



- If, after retesting of the initially reactive specimens, both wells are negative results (A/C.O.<0.9), these specimens should be considered as non-repeatable positive (or false positive) and recorded as negative. As with many very sensitive ELISA assays, false positive results can occur due to the several reasons, most of which are connected with, but not limited to, inadequate washing step. For more information regarding Wantai ELISA Troubleshooting, please refer to Wantai's "ELISAs and Troubleshooting Guide".
- If after retesting in duplicate, one or both wells are positive results, the final result from this ELISA test should be recorded as repeatedly reactive. Repeatedly reactive specimens could be considered positive for HEV IgG antibodies and therefore the patient is probably infected with HEV.
- After retesting in duplicate, specimens with values close to the Cut-off value should be interpreted with caution and considered as "borderline" zone specimen, or uninterpretable for the time of testing.

PERFORMANCE CHARACTERISTICS

1. Detection of HEV antibodies in specimens from patients with 10 years of HEV post infection history:

Reagents	Specimens	Pos. rate%	Cut-off	Positive specimens OD			Avr. pos S/CO
				lowest	avr.	highest	
WT IgG*	50	86	0.148	0.532	1.368	2.327	9.24
EIA 1**	50	36	0.512	0.514	0.018	2.415	1.98
EIA 2**	50	30	0.228	0.229	0.457	1.094	2.08

* WANTAI HEV-IgG ELISA

** Commercially available HEV IgG ELISA tests.

2. Detection of serial serums specimens from acute HEV phrase

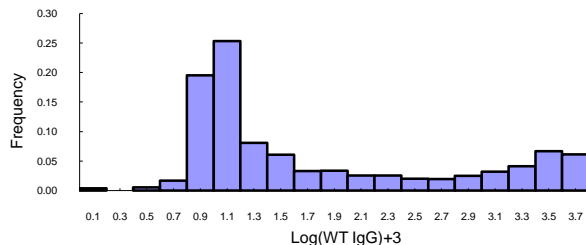
Since no golden standard for hepatitis E is available, the reference HEV IgG assay served as control reagent. Parallel comparison testing was performed with acute hepatitis E specimens (Testing Center 1). If the true positive status was defined as positive for any of the two tests used in this study, the sensitivity of Wantai HEV IgG and the reference HEV IgG was 97.96% and 91.84% respectively.

In the evaluation of 120 serial sera specimen obtained from 30 hepatitis E (Testing Center 2). The sensitivity of Wantai HEV-IgG were 100%, while the sensitivity of the reference HEV-IgG were 93.33%.

The sensitivity of Wantai HEV IgG was 99.08% in the parallel testing of a total of 218 serum specimens of acute hepatitis E, which was significantly higher than the reference HEV IgG tests (92.66%).

Testing Center	Case Number	WT-IgG		EIA 1**	
		Positive Number	Positive Rate	Positive Number	Positive Rate
1	98	96	97.96%	90	91.84%
2	120	120	100.0%	112	93.33%
TOTAL	218	216	99.08%	202	92.66%

3. Total 10587 blood specimens from blood donors were tested, the results are showed in the frequency distribution map, it exists two peaks of WT IgG: the first peak was higher, of which the center was near the OD value 0.0126, representing the people who did not infect HEV. The OD logarithm of the first peak was similar to the log-normal distribution. Considering the first peak as the center, calculate the standard deviation of the data on the left. The corresponding OD values in accordance with 99%-fractile and 99.9%-fractile respectively were 0.069 and 0.120 and the probability value at the cutoff value 0.185 was 99.99%. The second peak concentrated near the OD value 3.2, representing the HEV infection population, and its logarithm behaved as negative skewed distribution. As a result, the specificity of WT IgG was much higher, with false positive rate at 0.01%.



LIMITATIONS

1. Positive results must be confirmed with another available method and interpreted in conjunction with the patient clinical information.
2. Antibodies may be undetectable during the early stage of the disease and in some immunosuppressed individuals. Therefore, negative results obtained with WANTAI HEV-IgG ELISA are only indication that the specimen does not contain detectable level of HEV IgG antibodies and any negative result should not be considered as conclusive evidence that the individual is not infected with HEV.
3. If, after retesting of the initially reactive specimens, the assay results are negative, these specimens should be considered as non-repeatable (false positive) and interpreted as negative. As with many very sensitive ELISA assays, false positive results can occur due to the several reasons, most of which are related but not limited to inadequate washing step. For more information regarding Wantai ELISA Troubleshooting, please refer to Wantai's "ELISAs and Troubleshooting Guide", or contact Wantai technical support for further assistance.
4. The most common assay mistakes are: using kits beyond the expiry date, bad washing procedures, contaminated reagents, incorrect assay procedure steps, insufficient aspiration during washing, failure to add specimens or reagents, improper operation with the laboratory equipment, timing errors, the use of highly hemolyzed specimens or specimens containing fibrin, incompletely clotted serum specimens.
5. The prevalence of the marker will affect the assay's predictive values.
6. This kit is intended ONLY for testing of individual serum or plasma specimens. Do not use it for testing of cadaver specimens, saliva, urine or other body fluids, or pooled (mixed) blood.
7. This kit is a qualitative assay and the results cannot be used to measure antibody concentration.

REFERENCES

1. Reyes GR, Purdy MA, Kim JP, et al. Isolation of a cDNA from the virus responsible for enterically transmitted non-A, non-B hepatitis. Science 1990; 247: 1335–1339.
2. Clayson E, Innis B, Myint K, et al. Detection of hepatitis E virus infections among domestic swine in the Kathmandu Valley of Nepal. Am J Trop Med Hyg. 1995, 53:228–232.
3. Meng XJ, Purcell RH, Halbur PG, et al. A novel virus in swine is closely related to the human hepatitis E virus. Proc Natl Acad Sci USA, 1997, 94: 9860–9865.
4. Tei S, Kitajima N, Takahashi K, et al. Zoonotic transmission of hepatitis E virus from deer to human beings. Lancet 2003; 362(9381):371.
5. Zheng YJ, Zhang J, Xia NS. A debate about that hepatitis E is a zoonosis. Chinese J Zoonosis (in press).
6. Wang YC, Zhang HY, Xia NS, et al. Prevalence, Isolation, and Partial Sequence Analysis of Hepatitis E Virus From Domestic Animals in China. J Med Virol 2002, 67:516–521.

SUMMARY OF THE MAJOR COMPONENTS OF THE KIT:

Use this summary only as a reference and always follow the comprehensive method sheet when performing the assay. Note: the components of individual kits are not lot- interchangeable.

1. Microwell plate	Code 5	one
2. Negative Control	Code 8	1x0.5ml
3. Positive Control	Code 7	1x0.5ml
4. HRP-Conjugate	Code 6	1x12ml
5. Specimen Diluent	Code 9	1x12ml
6. Wash Buffer	Code 1	1x50ml
7. Chromogen Solution A	Code 2	1x7ml
8. Chromogen Solution B	Code 3	1x7ml
9. Stop Solution	Code 4	1x7ml

SUMMARY OF THE ASSAY PROCEDURE:

Use this summary only as a reference and always follow the detailed method sheet when performing the assay.

Add Specimen Diluent	100µl
Add Specimen	10µl
Incubate	30 minutes
Wash	5 times
Add HPR-Conjugate	100µl
Incubate	30 minutes
Wash	5 times
Coloring	50µl A + 50µl B
Incubate	15 minutes
Stop the reaction	50µl stop solution
Read the absorbance	450nm or 450/600–650nm

EXAMPLE SCHEME OF CONTROLS / SPECIMENS DISPENSING:

	1	2	3	4	5	6	7	8	9	10	11	12
A	Blank	S3										
B	Neg.	...										
C	Neg.	...										
D	Neg.											
E	Pos.											
F	Pos.											
G	S1											
H	S2											

CE MARKING SYMBOLS:

	In Vitro Diagnostic Medical Device		+2°C~+8°C Storage Conditions
	Use By		Batch
	Content Sufficient For <n> Tests		Instructions For Use
	CE Marking – IVDD 98/79/EC		EU Authorized Representative
	Catalog Number		Manufacturer
	Beijing Wantai Biological Pharmacy Enterprise Co., Ltd. No.31 Kexueyuan Road, Changping District, Beijing 102206, China Tel: +86-10-59528888, Fax: +86-10-89705849 Website: www.ystwt.com Email: wtexport@ystwt.com		

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